Exercise-induced cardiac troponin elevation: An update on the evidence, mechanism and implications

Polly Baker a,b,⁎, Todd Leckie b,c, Derek Harrington d, Alan Richardson a,b

a Centre of Sport and Exercise Medicine (SESAME), University of Brighton, UK
b Brighton Marathon Research Group, Brighton, UK
c Anaesthetics Department, Eastbourne DGH, East Sussex Healthcare Trust, UK
d Cardiology Department, Tunbridge Wells Hospital, Pembury Hospital, TN2 4QJ, UK

1. Introduction

It is well accepted that short bouts of moderate intensity exercise taken regularly are beneficial for health [1,2]. What is unclear is whether this still applies to those participating in strenuous and/or prolonged exercise. Some of this uncertainty has arisen from the growing literature demonstrating a rise in the cardiac biomarker, troponin (cTn), following endurance exercise. Understanding the significance of this is important. Firstly, to enable clinicians to give informed advice to those wishing to participate in such exercise and secondly to facilitate the interpretation of troponin levels in the context of an endurance event.

2. cTn testing in the clinical setting

Due to their ability to detect cTn much earlier, high-sensitivity cTn (HS-cTn) assays have largely replaced those of standard sensitivity in the clinical setting [3–5]. The enhanced sensitivity of these newer assays has led to the detection of cTn in healthy individuals [6] which in conjunction with the existence of biological and analytical variability [7,8] has made it harder to differentiate between a pathological and normal cTn value. To overcome this serial blood testing and the evaluation of cTn kinetics has become a fundamental component of the clinical assessment of chest pain patients.

The change criteria for a pathological rise between the two blood sampling points is assay specific and depends on a variety of factors including the timing of baseline sampling and the onset of symptoms. The key is that any change detected is greater than the combined biological and analytical variation. A 20% or greater change from an elevated cTn value is set as the threshold for diagnosis of myocardial necrosis [9,10] and represents a significant >3 standard deviations of variation associated with an elevated baseline concentration change in cTn on the basis of a 5–7% analytical total CV [10]. For clinical situations where the baseline sampling value is below the URL a change in the range of 50–60% is needed [11]. This is not error proof and thus it is recommended [12] that if the clinical
situation is ambiguous and the pre-test likelihood of disease high, additional sampling is performed.

The fourth definition of myocardial infarction includes serial testing as a criterion for its diagnosis [13]. However, it defines this detection of a rise and/or fall of cTn with at least 1 value above the 99th percentile upper reference limit (URL) as acute myocardial injury (Mln) and explains that this may exist as a clinical entity in its own right. In addition, this latest consensus statement updates the definitions of the five subtypes of MI. Criteria required for the diagnosis of types 1, 2 and 3 includes acute MI in conjunction with clinical evidence of myocardial ischaemia. The different subtypes are then differentiated by the aetiology of the myocardial ischaemia. MI caused by atherothrombotic coronary artery disease (CAD) and usually precipitated by atherosclerotic plaque disruption (rupture or erosion) is designated as type 1 MI. Evidence of an imbalance between myocardial oxygen supply and demand unrelated to acute atherothrombosis meets the criteria for type 2 MI. Cardiac death in patients with symptoms suggestive of myocardial ischaemia and presumed new ischaemic ECG changes before cTn values become available or abnormal meets criteria for type 3 MI.

The 2015 ESC guidelines for the management of acute coronary syndromes (ACS) in patients presenting without persistent ST-segment elevation [12] continue to recommend serial cTn testing and presents a 0 h/3 h and 0 h/1 h rule in and rule out algorithm for clinical use. Those patients presenting post-exercise typically fall within this cohort and thus utilisation of their suggested cTn testing schedule is logical.

3. cTn elevation following exercise

Earlier investigations using 2nd and 3rd generation assays reported significant elevations of cTnT and 1 following endurance exercise [14–18]. Deriving common findings from these studies has been challenging for several reasons. Variables, including exercise modality, intensity and duration of exercise, are not standardised between studies. Furthermore, the use of different assays, each with its own sensitivity and specificity and threshold levels for detection and limit, makes cross comparison of results difficult. Finally, the timing for sampling troponin can differ between trials. As described later, the levels of troponin rise and fall over 24 h making it imperative to standardise sampling time points.

By taking advantage of the existence of only one cTnT assay (second or third- generation) (Roche Diagnostics, Lewes, UK) and limiting the search criteria to cTnT, Shave and colleagues were able to combine data sets and overcome some of these limitations. The authors examined 1120 cases from 26 trials investigating cTn elevation related to marathon running and included the detection of cTn prior to the race, collection of plasma at 0 h/3 h and 0 h/1 h rule in and rule out algorithm for clinical use. Those patients presenting post-exercise typically fall within this cohort and thus utilisation of their suggested cTn testing schedule is logical.

find that all participants demonstrated a detectable HS-cTn following completion of the race and that 69.8% (296/424) had a value above the 99th percentile, the cut-off used for myocardial infarction. More recently Richardson et al. [20] evaluate HS-cTn using a fifth generation high-sensitivity assay (Elecsys, Roche Modular E170) and find that all 52 marathon runners have an increase above the reference value, reporting mean post-race values of 74 ± 30 ng/L. These studies do not differentiate between those elevated troponin levels of male and female participants. Although differences between the threshold set for the 99th percentile has been seen [21,22], evidence suggests that a change to differentiate between gender is not needed for diagnostic performance [23].

Enhanced sensitivity of assays has increased the number of detectable cTn values, including those above the level used to diagnose myocardial necrosis. Taking into account the central role troponin plays in the risk stratification of ACS it is imperative that the mechanism and thus significance of troponin rise is understood.

4. Mechanism of cTn release

Several theories have been proposed to explain the mechanism underlying Tn release following exercise. Currently the most well received is that of increased membrane permeability of cardiomyocytes, whereby unbound cTn found in the cytosol diffuses across a concentration gradient from the intra- to extra-cellular compartment. The initial peak, illustrated in Figure 2, would represent the release of Tn through the sarcoplasmic membrane with levels subsequently decreasing over 24 h reflecting the half-life and clearance of cTn subunits thereafter.

It has been suggested that mechanical stress through the transient disruption (wounding) of the sarcolemma might be responsible for this increased membrane permeability. Initially demonstrated in the skeletal muscle of rats after a period of downhill running [24], it was shown that cell 'wounds' resolve within 24 h following the same time-course as the CK levels detected. These events have also been demonstrated in-vivo cardiac muscle and have been associated with the release of growth factors [25,26]. It is logical to think, therefore, that troponin elevation following exercise may be a reflection of the adaptive cellular cascades seen in exercise-induced cardiac remodelling and hypertrophy.

An alternative hypothesis is that of integrin-stimulated troponin release. Integrins are transmembrane glycoproteins that mediate the attachment of cells to the extracellular matrix [27]. They are responsive to stretch, such as that seen in haemodynamic overload, and transmit force across the plasma membrane. This has been shown to trigger intracellular signalling pathways responsible for cardiac hypertrophy [28] and it is proposed that Tn is released as a by-product of this process. Recently, Hessel et al. [29] linked the stimulation of integrin to the reversible release of Tn from cultured cardiomyocytes. The pentapeptide Gly-Arg-Gly-Asp-Ser is a known agonist for integrin stimulation. Treatment of myocytes with this peptide resulted in a two to three-fold increase in Tn release versus various controls. Reversible myocardial injury was inferred by the absence of LDH release in treated cells relative to controls.

There is uncertainty as to how the intracellular cTn moves into the bloodstream. An alternative mechanism of increased membrane permeability has been suggested. This involves the formation of blebs which bud off from the plasma membrane of the cell, these have been seen in the liver in response to cellular ischaemia [30]. If the ischaemia is limited and re-oxygenation occurs the blebs may be released into the circulation without rupture of the plasma membrane, resulting in a one-off release of cytoplasmic contents. If the ischaemia is sustained the blebs will grow and eventually rupture leading to cell necrosis. Support for this process occurring in the heart comes from studies showing the presence of blebs on cultured cardiomyocytes subjected to hypoxia and release of cTn, without the development of cellular necrosis [31].
## Table 1
Overview of trials showing cTn release kinetics.

<table>
<thead>
<tr>
<th>Study (year)</th>
<th>N</th>
<th>Male/ female</th>
<th>Mean age (years)</th>
<th>Type of race</th>
<th>Troponin</th>
<th>Timing of sample collection</th>
<th>Highest prevalence of post-exercise cTn elevation</th>
</tr>
</thead>
<tbody>
<tr>
<td>Neumayr et al (2001) [15]</td>
<td>38</td>
<td>38/ 0</td>
<td>35</td>
<td>230km mountain cycle at 5500km altitude Marathon</td>
<td>TnI</td>
<td>Baseline, immediately and 1 day after the race</td>
<td>13/38 (34%) above URL of 0.05ug/L</td>
</tr>
<tr>
<td>Hermann et al (2003) [46]</td>
<td>46</td>
<td>40/ 6</td>
<td>40</td>
<td>Marathon</td>
<td>TnT TnI</td>
<td>Baseline, &lt;15 mins, 3 and 24 hours (9/46) after the race</td>
<td>TnT – not documented</td>
</tr>
<tr>
<td>Frassl et al (2008) [42]</td>
<td>15</td>
<td>0/ 15</td>
<td>37</td>
<td>Marathon</td>
<td>TnT TnI</td>
<td>Baseline, immediately and 24 and 72 hours after the race</td>
<td>TnT – 8/15 (53%) above URL/AMI cut off of 0.01ng/mL; TnI – 5/15 (33%) above URL of 0.03ug/mL; 15/26 (58%) above URL of 0.16ug/L</td>
</tr>
<tr>
<td>La Gerche et al (2008) [38]</td>
<td>27</td>
<td>20/ 7</td>
<td>32</td>
<td>Ultra endurance triathlon (3.8km swim, 180km cycle, 42.2km run)</td>
<td>TnI</td>
<td>Baseline, immediately and 1 week after the race</td>
<td>33/46 (72%) above URL of 0.04ug/L and 27/46 (58%) above AMI cut off of 0.06ug/L</td>
</tr>
<tr>
<td>Middleton et al (2008) [37]</td>
<td>9</td>
<td>9/ 0</td>
<td>-</td>
<td>Treadmill marathon</td>
<td>TnT</td>
<td>Baseline and then every 30 mins within the race and 1, 3, 6, 12 and 24 hours after race completion</td>
<td>9/9 (100%) within race and 8/9 (89%) after race completion, URL not documented 0/19 (0%) above URL of 0.04ug/L</td>
</tr>
<tr>
<td>Serrano-Osteriz et al (2011) [47]</td>
<td>21</td>
<td>19/ 2</td>
<td>38</td>
<td>Treadmill running for 45, 90 &amp; 180 mins at 85 and 90% IAT</td>
<td>TnI</td>
<td>Baseline, immediately after and 1, 3, 6, 12 and 24 hours after the race</td>
<td>91/102 (89%) above URL of 14ng/L</td>
</tr>
<tr>
<td>Nie et al (2011) [39]</td>
<td>12</td>
<td>12/ 0 adolescents</td>
<td>16</td>
<td>Track running</td>
<td>HS-TnT TnT</td>
<td>Baseline, immediately after and 2, 4 and 24 hours after the race</td>
<td>91/102 (89%) above URL of 14ng/L</td>
</tr>
<tr>
<td>Tian et al (2012) [45]</td>
<td>26</td>
<td>13/ 0 (adults) and 13/ 0 (adolescents)</td>
<td>24 and 14 respectively 34 9</td>
<td>90 minutes treadmill running at 90% VT Marathon</td>
<td>HS-TnT TnT</td>
<td>Baseline, immediately after and 1, 2, 3, 4, 5, 6, and 24 hours after the race</td>
<td>41/66 (62%) above URL of 14ng/L</td>
</tr>
<tr>
<td>Wilhelm et al (2012) [44]</td>
<td>10</td>
<td>10/ 0</td>
<td>39</td>
<td>Marathon</td>
<td>HS-TnT</td>
<td>Baseline, immediately after and 1, 5 and 8 days after the race</td>
<td>41/66 (62%) above URL of 14ng/L</td>
</tr>
<tr>
<td>Roca (2017) [43]</td>
<td>79</td>
<td>57/ 22</td>
<td>39</td>
<td>Marathon</td>
<td>HS-TnT</td>
<td>Baseline, immediately after and 1, 3, 6, 12 and 24 hours after the race</td>
<td>79/79 (100%) above URL of 14ng/L</td>
</tr>
<tr>
<td>Legaz-Arrese (2017) [40]</td>
<td>66</td>
<td>7/ 9 (adults) and 57/ 25 (adolescents)</td>
<td>31 and 15 respectively 40</td>
<td>Swimming</td>
<td>HS-TnT</td>
<td>Baseline, immediately after and 3, 6 and 24 hours after the race</td>
<td>41/66 (62%) above URL of 14ng/L</td>
</tr>
<tr>
<td>Baker et al (2019) unpublished</td>
<td>26</td>
<td>18/ 8</td>
<td>40</td>
<td>Marathon</td>
<td>HS-TnT</td>
<td>Baseline, immediately after and 3, 6 and 24 hours after the race</td>
<td>26/26 (100%) above URL of 14ng/L</td>
</tr>
</tbody>
</table>

BNP = brain natriuretic protein; TnI = cardiac troponin I; TnT = cardiac troponin T; CK = creatine kinase; h-FABP = heart-type fatty acid binding protein; HS-CRP = high-sensitivity C reactive protein; HS-TnT high-sensitivity troponin T; HBD = hydroxybutyrate dehydrogenase; IL-6 = interleukin 6; NT-proBNP = N terminal pro-hormone of brain natriuretic peptide; N = number of runners. This table illustrates all studies demonstrating cTn kinetics after exercise. It describes the basic demographics of the participants (age, sex), exercise performed and sampling points used.
Whether the release of cTn from the myocardium is related to intact subunits or degradation products is also under debate. In contrast to Hessel’s study [29] where intact cTn is released, Feng and colleagues [32] demonstrated the release of degraded cTn subunits in response to preload. Without having assays specific to cTn degradation products, the form of cTn released following exercise is not known.

Tn release following prolonged and/or strenuous exercise could be secondary to subclinical apoptosis or necrosis. The kinetics of exercise-induced cTn rise with a peak within the first 1–4 h and falling levels thereafter with resolution at 72 h make this suggestion unlikely. This in contrast to the peak seen at 24 h after a type 1 MI peak with continued sustained levels for several days [33]. Finally injured skeletal muscle may release proteins that are detected by TnT assays resulting in situations where elevated cTn values may originate from skeletal muscle [34–36]. This as yet has only been seen in those neuromuscular conditions and its applicability to healthy participants completing a bout of exercise is uncertain.

5. Kinetics of cTn release

In order to understand the underlying mechanism, it is useful to look at the kinetics of cTn release following a bout of endurance exercise. A review of the literature identifies 12 studies investigating the kinetics of cTn release following prolonged exercise [15,37–47] (see Table 1). All cTn kinetic trials except that by Middleton et al. [37] demonstrate a peak within the first 24 h after exercise. To establish the timing of the cTn peak blood samples must be drawn at multiple time points within this time. The most comprehensive data set is provided by Tian et al. [45] who conduct HS-cTnT sampling at 1, 2, 3, 4, 5, 6 and 24 h. They confirm a cTn peak at 3–4 h in 13 adolescents and 13 adults after 90 min of treadmill running at 90% of ventilatory threshold. However when evaluating cTn kinetic studies conducted following a marathon using the gold standard HS-cTnT assay [41,43,44] sampling is only performed from 24 h onwards and thus a comprehensive trend of the rise and fall of troponin cannot be seen. To expand on our understanding of cTn kinetics in the field we evaluated runners at multiple time points within the first 24 h of marathon completion. We believe we are the first study to do this and thus are providing valuable new information in this field.

6. cTn kinetics data following 2017 Brighton Marathon

As part of a subgroup analysis looking at cTnT rise following completion of the 2017 Brighton Marathon, 28 runners consented to further blood draws. Sampling timing included baseline, 0, 3, 6 and 24 h after the marathon. The HS-cTnT assay used (Roche Modular E170 (fifth generation); Basel, Switzerland) had a lowest limit of detection (LLD) of 3 ng/L and diagnostic levels for myocardial necrosis, based on the 99th percentile of the individual cTnT assay with a coefficient of variation ≤10%, were 14 ng/L. Baseline characteristics are given in Table 1.

All samples, including those taken before the race, were detectable for troponin with the HS-cTnT assay. Immediately following and 3 h after the marathon troponin levels for all participants exceeded the threshold for diagnosis of myocardial infarction. At 6 h this fell to 90% and at 24 h to 22% of participants (Fig. 1).

When comparing our data to that seen in the study by Tian et al. several differences are seen. Firstly, the timing of the cTn peak is earlier in our study with a maximal cTn value occurring at 1 or 3 h, with the majority occurring within the first hour after the marathon (17/26). Furthermore, data within Tian’s study demonstrates reduced variance which may be related to the matching of training status in participants and the exposure to an exercise regime with a shorter fixed duration and intensity. This is in contrast to our study where runners would set their own running pace and be exposed to additional environmental factors such as temperature. The disparity between the timing of peak cTnT is more likely to be related duration rather exercise intensity. Richardson et al. [20] showed that marathon runners exercised at 101 ± 5% of ventilatory threshold, the findings also demonstrated that working at a greater relative exercise intensity was associated with greater cTn. This intensity is much greater than that used by Tian et al. [45] or by Serrano-Ostariz et al. [47] who also demonstrated peak values to occur earlier after lower intensity work. Data showed working at 85% of the individual anaerobic threshold (IAT) induced peak values 30 min post exercise in comparison to 3 h for 95% of IAT. Therefore, the duration of exercise (265 ± 52 min in this study) seems to be the most important comparative factor, allowing Tn levels longer to rise before post-race measurement can occur.

Our data, and that of the studies found above, is at odds with the study by Middleton et al. [37]. They investigated nine trained males over the duration of a treadmill marathon. Although the data showed a cTn rise during marathon running, cTn then returned to baseline.

![Fig. 1. CnTnT kinetics pre-marathon and 0, 3, 6 and 24 h post marathon. Mean (X) and standard deviation of high-sensitivity cTnT values at baseline and 0, 3, 6 and 24 h. The dotted line indicates reference value (99th percentile) of 14 ng/L.](https://doi.org/10.1016/j.ijcha.2019.03.001)
before the end of activity. Additionally, participants demonstrated a bi-
phasic response with peak values returning 12–24 h post-race. This re-
sponse is difficult to explain but could be as a result of participants
recovering and attempting to restart daily activity 12–24 h post mara-
thon. The lack of in-exercise blood sampling in the present study and
the literature base is a limitation and requires further investigation.

7. Clinical approach to patients with post-exercise troponin elevation

CTn testing continues to play a central role in the risk stratifi-
cation and subsequent management of those with an acute coronary syn-
drome. Updated guidance on the diagnosis and management of MI
and Non ST-elevation ACS recommends the use of the 5th generation
HS-cTn assays [9,12]. Serial testing, of at least two sampling points, is
recommended to overcome the analytical and biological variation that
is associated with HS-cTn assays. Using our data, and that from other ki-
netics studies, we support the use of repeated samples to establish cTn
kinetics when attempting to differentiate between cTn elevation related
to exercise and that of other causes. Using this approach in combination
with the latest ESC guidelines for the management of acute coronary
syndrome without persistent ST-elevation [12] allows the formulation
of the below algorithm (Fig. 2) which updates that proposed by Shave
et al. [48]. It is important to note that the flow chart is no replacement
for clinical judgement and is meant to facilitate the clinical in their deci-
sion making process. We would urge, in the situation of clinical uncer-
tainty, that repeated cTn testing and further investigations including
evaluation by an experienced cardiologist occur in the hospital setting.

8. Further studies

Although a variety of plausible mechanisms for post-exercise cTn el-
evation have been proposed no one has been confirmed. Without
understanding the mechanism, elucidating the clinical significance is
difficult. With a large body of evidence indicating that cTn elevation in
other clinical situations has a quantitative negative prognosis the need
to establish this information has never been greater. Currently cTn ele-
vation following exercise is considered benign and affected individuals
are not counselled nor treated. Further investigations studying the
peak cTn value and/or the kinetics and its relation to future medical
events are warranted to ensure this line of action is appropriate.

9. Conclusion

CTn testing continues to be central to the assessment of patients
with a suspected ACS. The current use of HS-cTn assays reveals cTn ele-
vation in all those participating in exercise. In the short term these indi-
viduals show no increased risk of cardiac events however long term
data is lacking. Our findings and that from other studies suggest an
early cTn peak and that time frame therefore should be considered
when individuals present with elevated values following exercise. Tak-
ing into account the kinetics of post-exercise cTn elevation and the char-
acteristics of HS-cTn assays we propose a new updated clinical
algorithm for the management of those presenting with clinical symp-
toms compatible with a cardiac event after exercise.

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